Urol Res (2001) 29: 152–162 © Springer-Verlag 2001

### INVITED EDITORIAL

D. Robert Siemens · Timothy L. Ratliff

# **Vaccines in urologic malignancies**

Received: 28 October 2000 / Accepted: 9 February 2001

**Abstract** The prospect of activating the immune system to combat neoplastic disease has stimulated the interest of clinicians and scientists for over 100 years. Despite a few notable exceptions (especially with urologic malignancies), immunotherapy has not fully reached its considerable therapeutic potential for the treatment of cancer. Tumors undoubtedly express antigens that may act as targets for antitumor immunity, and advances in molecular biology and tumor immunology have recently revived the possibility of a cancer vaccine. This improved understanding has resulted in numerous successes with active immunotherapy in animal models and has facilitated the clinical testing of cancer vaccines. Ongoing advances in the identification of unique, tumor-specific antigens and their presentation to stimulate T cells will be necessary for the emergence of these novel vaccine therapies for cancer patients. Herein we review the current concepts of tumor immunology, including observations on cell types probably involved with the immune surveillance of tumors, the presentation and recognition of "foreign" antigens, and possible mechanisms of tumor escape from the immune response, all of which are critical to the understanding of new initiatives for cancer vaccine therapy. Finally, we review some of the successes and limitations of vaccine therapy for urologic malignancies.

**Key words** Immunotherapy · Genitourinary neoplasms · Tumor immunity

D. R. Siemens (⋈)

Department of Urology, Queen's University, Kingston, 76 Stuart Street, Kingston, Ontario K7L 2V7, Canada Tel.: +1-613-5482411; Fax: +1-613-5451970 e-mail: siemensr@kgh.kari.net

T. L. Ratliff University of Iowa Department of Urology, Iowa City, Iowa, USA

# Introduction

The ability to distinguish self from nonself is the hallmark of the immune system and facilitates our survival in an inordinately hostile environment. The human immune system not only allows the appropriate recognition and destruction of nonself but also the ability to recognize and tolerate self. Classically, vaccines have taken advantage of this phenomenon by stimulating a humoral response to specific bacterial or viral antigens to prevent disease caused by these infectious agents. The appeal of a vaccine for neoplastic cells is enormous and has been stimulated by immune surveillance theories whereby newly transformed cells are detected and eliminated by an intact immune response. Indeed, the study of tumor immunology is based on two simple propositions: (1) tumor cells express distinct antigens that are found in only negligible amounts in normal cells and (2) these antigens can be recognized by the immune system, leading to destruction of the "foreign" cell as though it were a transplant allograft or infectious agent.

Since the first description of the presence of tumorassociated antigens, numerous distinctive cell surface molecules have been identified that could be recognized as "foreign" by the host immune system. It therefore seems that for a tumor to establish itself and eventually to metastasize, it must first escape the surveillance of immune cells percolating throughout virtually all tissues of the body. Why surveillance of the immune system against tumors is not more effective is relatively poorly delineated, and investigation of these concepts has contributed immensely to our understanding of tumor development and may have important implications for cancer management. Several investigators interested in urologic malignancies including renal cell carcinoma and prostate cancer pioneered many of these early investigations. Subsequently, several tumor-associated antigens were identified for urologic cancers, although most of these targeted antigens are also found on normal cells (gp100, PSA, PSMA, etc.) or are developmental antigens. Few truly foreign antigens have been identified.

# **Tumor immunology**

The goal of tumor immunology is to understand the immune response to malignant cells and create novel therapeutic strategies. In humans, the concept of tumor surveillance by the immune system is somewhat vindicated by the increased occurrence of some rare tumors with chronic immunosuppression. Additionally, in rare cases of melanoma or renal cell carcinoma, spontaneous regressions prove the ability of the immune system to reduce tumor progression [61]. Furthermore, studies have demonstrated a reduced cell-mediated immunity in some cancer patients, and it has been observed that reduced NK cell cytotoxic activity may play an important role in prostate cancer development [49] and metastases [88]. Although this impaired cellular immunity can be demonstrated in cancer patients (especially those with advanced disease), it is likely that a generalized immunologic deficit is not causative but instead reflects a secondary phenomenon. In fact, the vast majority of cancers do not increase in immunodeficient hosts, suggesting that the normal immune response is unable to control most forms of cancer.

In the early twentieth century, Coley used heat-killed bacterial infections to initiate an antitumor response [32]. His observations led to the supposition that tumor escape mechanisms could be overridden to induce an antitumor response. However, clinically effective treatment failed to materialize until the advent of bacille Calmette-Guérin (BCG) for the treatment of superficial bladder cancer [50]. Rosenberg and associates revitalized interest in immunotherapy with their work on lymphokine-activated killer cells and tumor-infiltrating lymphocytes [3, 95]. Although therapeutic efficacy fell short of expectations, these studies demonstrate the ability of immune cells to eliminate tumors previously thought resistant to immune effector mechanisms. Gene therapy studies have also confirmed that most theoretically "nonimmunogenic" tumors are indeed immunogenic [5, 20, 99]. Nonetheless, numerous early studies demonstrated that specific immune cells recognize tumor-specific antigens, with resultant lytic activity of the tumor cells [80, 91]. In fact, a vast number of potential antigens result from any of several genetic alterations in cancer cells, including any amino acid sequence in any membrane-bound or intracellular protein.

### Cell-mediated immunity

Since the discovery of tumor-specific transplantation antigens, cell-mediated immunity has been recognized as the predominant immune effector response in tumors, especially solid tumors. Cell populations involved in the immune response include lymphocytes, granulocytes, and specialized antigen-presenting cells (APC). The granulocytes include neutrophils which, along with macrophages and monocytes, are important for phagocytosis of antigens targeted by antibodies. The specialized APC include monocytes, macrophages, Langerhans' cells, Kupffer's cells, and dendritic cells. These cells express major histocompatibility complex (MHC) molecules classes I and II to display antigens appropriately to T lymphocytes (T cells). T cells, B lymphocytes (B cells), and natural killer (NK) cells make up the three major populations of lymphocytes and can be defined by the presence and type of transmembrane antigen receptors. The NK cells are large granular lymphocytes capable of killing certain cancer cells with no prior immunization and without restriction by MHC glycoproteins (which require T cells to recognize antigen in the context of self).

Although numerous cell types may be involved in antitumor immunity, including macrophages, antibody-dependent cell-mediated cytotoxicity (ADCC), and NK cells, evidence is emerging that T cells are most important in developing host antitumor activity. Mature T cells are comprised of two major populations defined by the cell surface expression of CD4 or CD8 molecules. The CD4+ cells recognize antigen when presented in association with MHC class II molecules, while CD8+ T cell (cytotoxic T cell or CTL) antigen recognition is restricted by MHC class I expression (Fig. 1).

T cells can also be classified by their functional role in modulating the immune response. Classically, CD4+cells (helper T cells) have been thought to play an assisting role in stimulating other cells such as B cells, other T cells, and macrophages, whereas the CD8+T cells are cytotoxic to cells displaying the antigen that they recognize. Furthermore, when naive T cells are activated, they produce interleukin-2 (IL-2) and then differentiate into two distinct subpopulations that can be categorized according to the different cytokines they produce [1].

The T helper subset,  $T_H1$  cells, secrete predominantly IL-2, IFN- $\gamma$ , and TNF- $\beta$  and promote cell-mediated immune responses (i.e., delayed-type hypersensitivity, or DTH). During antigen presentation, APC produce IL-12, which drives T cells to differentiate toward a  $T_H1$  response. IFN- $\gamma$  secretion by  $T_H1$  cells acts as an autocrine agonist and will inhibit differentiation of  $T_H2$  cells, another T helper subset.  $T_H2$  cells are the principle T helper cell for B cell function and propagation of the humoral allergic immune response. The cytokines they produce are predominantly IL-4, IL-5, IL-9, IL-10, and IL-13. IL-4 is known to enhance differentiation to a  $T_H2$  response and at the same time antagonize the  $T_H1$  response (Fig. 1).

The ability of the T cell to recognize antigen is a property of the T cell receptor (TCR) [11] and is specific for a certain peptide antigen/MHC combination. The TCR is a disulfide-linked heterodimer very similar in structure to the Fab fragment of an immunoglobulin molecule and is associated with signal-transducing

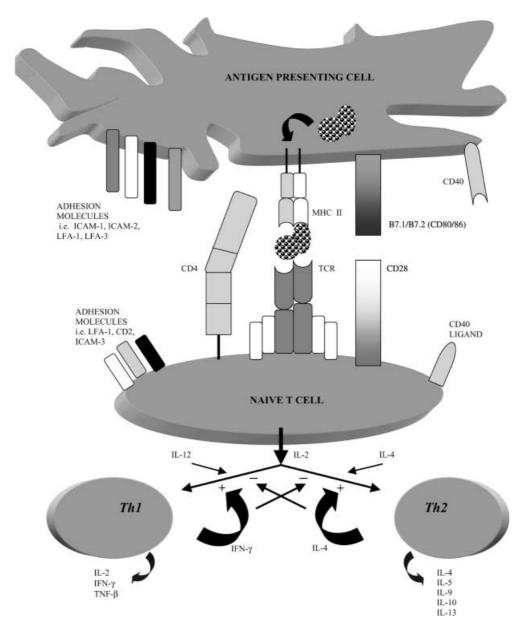


Fig. 1 Major histocompatability complex (MHC) class I molecules are found on virtually all nucleated cells and restrict antigen presentation to cytotoxic T cells. The MHC class II molecule is found only on the specialized antigen presenting cells such as macrophages, dendritic cells and B cells and restricts antigen presentation to helper T cells through the interaction with the CD4 molecule (as shown in the Fig. 1). Activation of T cells occurs after antigen presentation by the MHC molecules, and requires complex signaling by both the T cell receptor (TCR) and ligation of CD28 by the B7 molecules (CD80/CD86) on the APC. Another important costimulatory molecule is the CD40 molecule expressed on antigen presenting cells such as dendritic cells. Molecules necessary for cellular adhesion include LFA-1, CD2 and ICAM-3 on the T cell as well as ICAM-1, ICAM-2, LFA-1 and LFA-3 on the antigen presenting cells. Differentiation of T cells into T<sub>H</sub>1 and T<sub>H</sub>2 lymphocytes requires a complex interaction involving numerous immune cells and secreted cytokines, including IFN-y secretion for T<sub>H</sub>1 differentiation and IL-4 for T<sub>H</sub>2 differentiation. The subsets are characterized by their secreted cytokine profile: IL-2, IFN- $\gamma$  and TNF- $\beta$  for T<sub>H</sub>1 cells and IL-4, IL-5, IL-9, IL-10 and IL-13 for T<sub>H</sub>2 cells. T<sub>H</sub>1 subsets promote cellmediated immune responses whereas T<sub>H</sub>2 cells induce humoralmediated allergic inflammation

molecules known as the CD3 complex [35]. The extracellullar engagement of peptide antigen/MHC molecule to the TCR is insufficient to initiate the transduction of these intracellular signals. Important costimulatory molecules distinct from the TCR complex are also required for appropriate T cell activation. The most studied costimulatory signal is the ligation of a T cell surface molecule, CD28, to the CD80 (B7-1) or CD86 (B7-2) of the APC, as this is a requisite second signal for T cell activation [42]. Other cell surface molecules such as ICAM-3, LFA-1, and CD-2 are also important in T cell interactions with APC.

For T cells to recognize antigens and discriminate self from nonself, the antigenic peptides must be presented in association with major histocompatibility complex (MHC) molecules. The MHC is a genetic locus on chromosome 6 that encodes the cell surface structures, human leukocyte antigens (HLA) or H-2 in mouse.

There are two structural classes of HLA molecules, each of which restricts antigen recognition to different T cells. The three-dimensional structure of the class I and II molecules exposes a groove involving 8 to 25 amino acids that can be recognized by the TCR of the T cells. With the aid of the MHC molecules, T cells can recognize foreign antigens in the context of self, as the TCR recognizes a composite of antigens associated with the correct MHC molecule.

Class I HLA molecules are found on virtually all nucleated cells and restrict CTL. They are composed of a 44-kd  $\alpha$  chain encoded within the MHC. This glycoprotein is associated with  $\beta$ 2-microglobulin encoded by a non-HLA gene found on chromosome 15 [12]. HLA-A, -B, and -C are the three major class I genes and numerous alleles are defined for each of them [100]. Class II molecules are also heterodimeric structures, although they are found only on specialized antigen-presenting cells such as macrophages, dendritic cells, and B cells and restrict antigen recognition to T helper cells. There are also only three major categories of class II molecules, HLA-DR, -DQ, and -DP, and each molecule is made up of  $\alpha$  and  $\beta$  chains encoded within the MHC [14].

### Tumor escape mechanisms

Why the immune surveillance is not more effective in controlling tumor formation/progression is poorly understood. One possible mechanism of tumor escape is the selection of tumor cell clones expressing fewer immunodominant antigens by the pressure of normal host immune surveillance. Several studies have documented the outgrowth of tumor cell lines with few tumor-specific antigens, and this loss of antigen expression could be due to antibody-induced internalization or antigenic variation. Fortunately, even immunoselected tumor cell variants have been shown to express a number of unique antigens that could serve as targets for immunotherapy protocols [23].

Even if stable molecules are present, the tumorbearing host may become tolerant to these antigens. This tolerance may be due to improper antigen presentation or neonatal exposure to the antigen, as in the case of carcinoembryonic antigen (CEA). However, studies have demonstrated that immunization with some of these antigens can overcome tolerance and subsequently may be used for immunotherapy regimens [60].

Downregulation of the MHC molecules may also lead to tumor escape, as the lack of MHC input would result in ineffective presentation of antigen to immune cells in the context of self and may lead to suppression of the appropriate immune response. Downregulation of MHC class I molecules often increases tumorigenesis in animal models. Wallich showed that transfecting the MHC class I genes can inhibit the metastatic ability of tumor cells [105]. Bander et al. has confirmed lost or diminished class I expression in a number of cell lines as

well as in frozen tissue specimens of prostate cancer [7]. Levin demonstrated that less differentiated prostate tumors expressed significantly fewer MHC class I molecules and, when both class I expression and degree of differentiation were considered, those with higher expression had better survival [55]. The authors suggest that HLA class I status may be an important prognostic factor as well as an important target for future immunotherapy strategies.

As discussed earlier, effective CTL activation is dependent on stimulatory signals from the T<sub>H</sub>1 subpopulation of helper T cells. As most solid tumors do not express class II MHC molecules necessary for antigen presentation to the helper T cells, tumor development may take advantage of the lack of activating signals. Recent studies have revealed that in a number of advanced malignancies, the zeta ( $\zeta$ ) chain of the T cell receptor in tumor-infiltrating lymphocytes shows decreased expression [25] and that this loss is associated with poorer prognosis [111]. Healy similarly demonstrated impaired expression and function of  $\zeta$  chains in the peripheral blood lymphocytes in patients with advanced prostate cancer [36]. A number of molecules have been shown to be critical for effective activation of T cells, and the lack of these costimulatory signals may lead to tolerance of T lymphocyte responses. The best characterized of these costimulatory molecules is the B7-CD28 receptor ligand pair [54], which has been shown in animal studies to augment the antitumor immune response when amplified. Other molecules necessary for the binding of lymphocytes to antigen-presenting cells include LFA-1, CD2, and ICAM-3 on T cells as well as ICAM-1, ICAM-2, LFA-1, and LFA-3 on the antigenpresenting cells.

The resistance of certain populations of neoplastic cells to cytotoxic cell killing mechanisms may be another mechanism of tumor escape. The inability of tumor cells to undergo Fas-mediated death may contribute to evasion of immune surveillance. Lehmann et al. recently described a novel method of tumor escape to NK-mediated killing. They describe the impaired binding of perforin on the cell surface of tumor cells, with subsequent resistance of granule-mediated cell death from the cytotoxic effector cells [53].

Investigators have demonstrated that tumor cells can further affect their environment to potentially decrease the effectiveness of any host antitumor immune response. Examples include the release of free antigens that can interfere with the effective response of NK cells and helper T cells. Also, tumor cell production of cytokines such as IL-10 and IL-18 has been shown to limit the effectiveness of immune surveillance. Transforming growth factor- $\beta$  (TGF- $\beta$ ) has been demonstrated to inhibit IL-2-dependent immune responses of macrophages and T cells [26]. Another mechanism of tumor escape may involve Fas/Fas ligand interaction. Several human tumors have been found to express Fas ligand, which can induce Fas receptor-mediated apoptosis of activated immune cells [10]. Kim and associates found high

expression of Fas ligand and a relatively low expression of Fas in renal cell carcinoma, which may be involved in the evasion of immune effector cells, although no correlation could be found with tumor stage or grade [47].

Finally, it is possible that host cells can be recruited to the tumor site for active downregulation of antitumor T cell immune responses [74]. There is some experimental evidence for the existence of T cell subsets that may play an antigen-specific suppressor role in tumor immunology. These T cells have been shown to be CD4+ and may have T<sub>H</sub>2 characteristics, although their role in human cancers has been difficult to determine.

# **Tumor immunotherapy**

Numerous therapeutic strategies utilizing the immune system to eradicate neoplastic disease have been developed (Table 1). Early attempts focused on nonspecific strategies and the best example of cancer immunotherapy is still the in situ use of bacille Calmette-Guérin (BCG) for carcinoma of the bladder. Although the exact antitumor mechanism has still not been completely delineated, it undoubtedly relies on a functioning immune system [75] and, in particular, requires the presence of both CD4+ and CD8+ T cell subsets [76].

Adoptive immunotherapy is a passive, nonspecific technique involving the transfer of immunologically activated lymphoid cells. Clinical experience has revealed that the activation of human lymphoid cells, i.e., IL-2-activated LAK cells (TIL), is feasible and that systemic administration is safe; however, clinical success in protocols for renal cell carcinoma and melanoma has been limited [3, 95]. Lubaroff et al. [56] have shown that a severe combined immunodeficiency (SCID) mouse model is a viable system for studying adoptive therapies for human prostate cancer and, furthermore, demon-

strated antitumor activity utilizing autologous IL-2-activated tumor-infiltrating lymphocytes (personal communication). A novel approach to adoptive therapy was reported by Cesano et al. whereby the MHC non-restricted human T cell line TALL-104 demonstrated a significant antitumor effect against DU-145 tumors in SCID mice [16].

The systemic administration of cytokines such as IL-2, IL-4, and recently IL-12 is an example of active nonspecific immunotherapy. The first extensive studies of cytokine immunotherapy were performed with IL-2, and remissions reported in patients with renal cell carcinoma and melanoma give a "proof of principle" of tumor immunotherapy, even for metastatic disease. IL-12, a heterodimeric cytokine with immunoregulatory activity, has shown potent antitumor activity in a number of different murine tumor models. However, enthusiasm for these systemic cytokine therapies needs to be tempered by the frequent reports of severe toxic effects.

#### Cancer vaccines

An alternative immunotherapeutic strategy to simple systemic administration of cytokines or activated immune cell populations is generation of a vaccine that could elicit a specific antitumor response in vivo. In this regard, there are a number of significant differences in the development of vaccines against infectious agents as opposed to neoplastic cells. Viral genes, for example, are relatively simple, dictating a limited number of defined antigens to be manipulated for immunization. Tumor cells, on the other hand, possess a virtually unlimited number of antigens, the majority of which have not been identified. Vaccination against infectious agents occurs prior to exposure in order to prevent disease, whereas in the case of a cancer vaccine, the immune system has

Table 1 Immunotherapy strategies

Type	Description	Example
Passive immunotherapy		
Anti-tumor antibodies	Administration of tumor-specific antibodies Complexed with toxic materials, drugs, radioisotopes	Radioisotope-labeled anti-PSMA Anti-PSA carrying chemotherapeutics
Adoptive immunotherapy	Transfer of activated lymphoid cells	IL-2 activated LAK cells Human T cell line TALL-104
Active immunotherapy		
Cytokines	Systemic administration of cytokines to stimulate an antitumor response	IL-2 for renal cell carcinoma
		IL-4, IL-12
Tumor vaccines		
Antigen-specific	Recombinant peptide stimulating an antitumor response delivered as	PSA/PSA segment vaccine
	– DNA vaccine	Vaccinia virus/PSA vaccine
	<ul> <li>Peptide/protein vaccine</li> <li>Viral or bacterial vaccine</li> <li>Peptide-pulsed dendritic cell vaccine</li> </ul>	Dendritic cell/PSMA vaccine
Tumor cell vaccine	Ex vivo administration of modified autologous or allogeneic tumor cells	Autologous prostate tumor cell vaccine
	In situ modification of tumors to stimulate antitumor response	In situ transfer of cytokines to tumors, i.e., IL-2 and IL-12

theoretically already been exposed to the tumor antigens and therefore may already be "tolerant" to them. Finally, the majority of cancer vaccines focus on stimulating a different arm of the immune response, cell-mediated immunity, not the usually desired humoral response to infectious agents.

There are generally two ways of categorizing cancer vaccines, depending on the source of the immunizing antigens: antigen-specific or cell-based cancer vaccines [34].

# Antigen-specific vaccines

Antigen-specific vaccines involve the delivery of recombinant peptides or proteins to a host in order to elicit an antitumor immune response. Much interest has been stimulated in this possibility since the first convincing demonstration of the existence of specific tumor-associated antigens [72], and the development of several novel techniques to define MHC class I-restricted antigens molecularly has facilitated an explosion of research in this area [13, 15, 37, 40, 46, 66, 70, 79]. Any immunotherapy protocol has several different means of delivering these antigens to the host:

- 1. DNA-based vaccines
- 2. Peptide- or protein-based vaccines
- 3. Recombinant viral or bacterial vaccines
- 4. Antigen-pulsed dendritic cell vaccines

DNA vaccines encoding tumor antigens can be delivered encapsulated by liposomes or as naked DNA [66]. Despite the possibility of rapid degradation of DNA when given systemically, there has been renewed interest in DNA-based vaccines for both infectious agents and neoplastic diseases [96, 108]. Xiang et al. recently described an autologous oral DNA vaccine delivered with an attenuated strain of Salmonella typhimurium to immunize mice successfully in a self-antigen model of melanoma [108].

The advantages of administering a cancer vaccine as a peptide or protein would include greater safety than with other vector-based vaccines. Tumor antigens could be delivered simply as defined peptides or as proteins specifically designed to access the class I pathway of APC in vivo. Several phase I clinical trials have been completed utilizing peptide-based vaccines for malignant melanoma, demonstrating minimal toxicity and some clinical and in vitro evidence of response [58, 71, 83].

Advances in recombinant technology have stimulated interest in the incorporation of genes encoding relevant antigens into vectors such as bacteria or viruses to augment their immune response. Vectors such as *Listeria* monocytogenes, adenovirus, and the pox viruses (including vaccinia) impart a number advantages including high-efficiency gene transfer and targeting of the MHC class I and II antigen-processing pathways [39, 43, 69, 78, 112].

While viruses are effective in generating CTL and ultimately antitumor activity in naive mice, the use of

viruses in settings where the host has been previously exposed to the virus significantly reduces transgene expression [48, 109]. Studies by Yang and associates formally demonstrated that T cell responses, specifically of CTL, to viral proteins were responsible for destruction of cells expressing the transgene [109]. Furthermore, antibodies in the serum have been shown to reduce the ability of viruses to deliver the transgene [48], and the resulting decrease in gene expression reduces subsequent CTL activation [41, 63]. The diminished ability to generate CTL when antibodies to the viral vector are present has been suggested as an important reason for the lack of CTL after adenovirus delivery of melanoma antigens in clinical trials [84].

To facilitate the presentation of antigenic peptides as a cancer vaccine, much interest has focused on the use of dendritic cells. These bone marrow-derived cells have the ability to process antigens and present them in the context of MHC molecules and other costimulatory molecules such as B7. They are the most potent APC identified and are capable of activating naive T cells. A potentially powerful strategy in cancer gene therapy involves the genetic engineering of dendritic cells with defined tumor antigens and their use as vaccines [97]. Dendritic cell vaccines can be produced by loading these cells in vitro with peptides, proteins, and whole tumor cells or by infecting them ex vivo by viral vectors [94]. The first few clinical trials utilizing a dendritic cell-based vaccine for metastatic melanoma and follicular B-cell lymphoma have demonstrated immunologic and clinical response [38, 65].

# Tumor cell vaccines

Active immunization can also be achieved by the administration of autologous or allogenic tumor cells modified to secrete cytokines or other immunostimulatory molecules capable of recruiting antitumor effector cells. Examples include in-vitro tumor cell transfection by MHC class II genes [68], transfection of lost MHC class I alleles [105], and transfection of the costimulatory molecule B7 to stimulate T cells [8]. The strategy of using tumor cell vaccines as a source of tumor antigens eliminates the need to identify and develop immunodominant antigens. Engineered tumor cells expressing cytokines may function as well by providing necessary growth factors to activated CTL in the absence of helper T cells. Numerous different cytokines have been introduced into tumor cells in vitro, including IL-2, IL-4, IL-6, IL-12, TNF- $\alpha$ , and GM-CSF. Many of these preclinical models have shown great success in controlling local tumor outgrowth as well as some measure of systemic protection. Numerous phase I/II clinical trials utilizing ex vivo transduced whole-tumor cell vaccines (most often involving malignant melanoma) have been published [22, 29, 90].

Cancer gene therapy strategies involving the transfer of genetic material in situ by viral and nonviral vectors have also been successful in stimulating active antitumor immunization. These in vivo schemes offer a number of practical advantages over their ex vivo counterparts, including the cost reduction of bypassing the isolation of target cells from the patient. Putzer demonstrated impressive regression of preestablished tumors through the intratumoral injection of adenovirus recombinant for IL-12 and the B7-1 molecule [73]. We have also observed significant antitumor activity of murine prostate cancer nodules after intratumoral injection of the canarypox virus, ALVAC, recombinant for murine IL-2, IL-12, and TNF-α.

# Vaccines in urologic malignancies

Immunotherapy for urologic malignancies has a rich history, given its arguably gold-standard applications in bladder and renal cancer. Advances in molecular techniques and knowledge of the immune response to tumor cells have generated great enthusiasm in the urologic community to the possibility of novel alternative or adjuvant approaches to the neoplastic diseases that we treat.

#### Prostate cancer

Despite the enormous prevalence of prostate cancer, its optimal management remains at best controversial, with 5-year biochemical failure rates for radical prostatectomy ranging from 27% to 57%. It is imperative to develop alternative or adjuvant treatment strategies, and immunotherapy presents a unique opportunity for better management of clinically localized and metastatic disease.

Prostate cancer is truly an exciting model for immunotherapy trials. Not only do numerous animal models exist, but human prostate cancer has also shown a variety of tumor-associated antigens. A few examples of these are PSA, hK2, PSCA, PSMA [40], MUC-1 [15], MUC-2 [28], GLOBO-H [46], GM-2 [45], and Lewis Y [37], some of which may well serve as targets for antigenspecific immune recognition [93]. Segments of the PSA protein have been shown to be immunogenic and stimulate a specific CTL reaction when used as a vaccine. Several peptide antigens, two 10-mer peptides from PSA (PSA-1 and PSA-3), and a PSA oligoepitope peptide (PSA-OP), have also been shown to elicit CTL responses in vitro [21]. From this initial work, several phase I/II clinical studies are underway in which patients are vaccinated against the PSA protein with the hope that systemic immunity against prostate cancer will occur. Phase I clinical trials have also been performed using autologous dendritic cells associated with the HLA-2.1specific prostate-specific membrane antigen (PSMA). Little adverse effect was observed and several hormonerefractory patients showed significant responses to the vaccine [86]. The phase II trial using this dendritic cell vaccine demonstrated an overall response rate of 30% (19/62), with 11 durable responses [98].

Several investigators have developed novel tumor cell vaccine strategies to treat prostate cancer. Early studies by Sanda and associates showed that GM-CSF-transfected rat prostatic adenocarcinomas grew more slowly than parental tumors [87]. Subsequently, Vieweg showed that IL-2 transfected rat R3327-MatLyLu also induced antitumor activity [102]. Existing tumors had reduced rates of tumor outgrowth, and protection was gained against subsequent tumor challenge. Using the parental R3327G tumor that exhibits hormone responsiveness, Yoshimura also observed antitumor activity; however, neither CTL activity nor protection against subsequent tumor challenge was observed [110]. We reported one approach to cancer immunotherapy involving the transfer of genes encoding the cytokines IL-2 and TNFα using the canarypox viral vector, ALVAC [44]. The ALVAC virus was shown to infect efficiently murine prostate cancer cells, RM-1, and produce high levels of extrinsic gene product. Antitumor immunity was also induced when tumor cells were infected by ALVAC cytokine recombinants and injected subcutaneously in the flanks of male C57BL/6 mice. Based on the optimistic results of such preclinical studies, an autologous vaccine approach is being investigated in a phase I/II study at Johns Hopkins Clinic in patients with extracapsular disease following radical prostatectomy. An allogeneic vaccine using MHC class I-matched allogeneic cells transduced to secrete IL-2 and IFN-γ is being employed in a NCI phase I/II study at the Memorial Sloan Kettering Institute. Likewise, a phase I clinical protocol for the intratumoral (in situ) injection of IL-2 and GMCSF genes is underway for prostate cancer.

# Kidney cancer

Management of metastatic renal cell carcinoma (RCC) has provided a "proof of principle" for immunotherapeutic approaches to the treatment of cancer. Metastatic RCC is a refractory disease that typically responds poorly to other interventions including radiotherapy and chemotherapy. However, systemic administration of cytokines such as IL-2 and interferon-α resulted in response rates from 15% to 25% [82, 89, 104, 107]. Further evidence for the ability of RCC to act as targets for immunomodulatory techniques include the clinical response rates to adoptive immunotherapy utilizing lymphokine-activated killer cells and tumor-infiltrating lymphocytes [9, 24, 81]. Similarly, Childs et al. recently described graft-vs-tumor responses after nonmyeloablative allogenic stem cell transplantation [19].

A recently described tumor marker for renal cell carcinoma, G250 antigen, has potential as a target for an antigen-specific vaccine-based therapy. It is a transmembrane protein identical to the tumor-associated antigen MN/CAIX [33] and highly expressed on kidney cancer cells, with little expression in normal cells [6]. This antigen has been a target for immunohistochemical studies [4] as well as a radioimmunotherapeutic clinical

trial [106]. The use of G250 as a target for vaccine therapies has been investigated in several preclinical reports. Van Dijk et al. reported on the enhancement of an anti-tumor immune response in a xenogenic model of renal cell carcinoma using the administration of antibodies to G250 [101]. Subsequently, Vissers et al. have analyzed the immunogenicity of G250 epitopes and concluded that this antigen may well serve as an important target for anti-RCC immunotherapy [103].

Despite the identification of G250 as an organ-specific antigen, relatively few tumor-associated antigens have been described for RCC. Instead, numerous reports have looked at the potential of using whole tumor cells as a source of potential antigens [2, 27, 59, 77, 85]. These clinical studies employed autologous allogenic tumor cells injected with immune adjuvants such as BCG, C. parvum, and Candida antigen. Although specific conclusions are difficult to ascertain based on the marked differences in these trials, it is encouraging that evidence of some clinical tumor regression or immune activation was consistently observed. Galligioni et al. [27] initiated a clinical trial utilizing an autologous tumor cell vaccine in patients after radical nephrectomy. The tumor cells were injected intradermally with BCG as an adjuvant. Although a survival benefit was not observed, significant differences in the DTH response to the tumor cells compared to normal renal cells did suggest the presence of a tumor-specific cell-mediated immune response.

Numerous technical advances have facilitated our ability to improve the immunogenicity of tumor cells with the transfer of genes encoding various cytokines such as TNF, IL-2, IL-4, and GM-CSF as well as costimulatory molecules such as B7 [30, 31, 92]. In this regard, Gansbacher et al. [30] reported on a pilot study of an allogeneic vaccine secreting IL-2, and Simons et al. [92] demonstrated provocative results with an autologous GM-CSF-secreting vaccine for patients with metastatic renal cell carcinoma.

### Bladder cancer

Since the first instillation of BCG into the bladders of patients with superficial TCC [62], it has become increasingly clear that bladder cancer is the most immunosensitive tumor and an ideal candidate for future immunotherapy research. The observations confirming the role of an intact immune response in the mechanism of antitumor activity for BCG have stimulated several trials utilizing immunomodulating techniques. Some of these include the intravesical instillation of cytokines such as IL-2 and the interferons. O'Donnell et al. reported on the safety and efficacy of a modified BCG that expresses the gene for IL-2 [18, 67].

Although several tumor-associated proteins have been identified for bladder cancer, most were investigated as biologic markers for the detection of disease and not as possible targets for a vaccine strategy. However, Luo et al. identified a mutated form of ras p21 protein in the MB49 murine cell line, and immunization resulted in mutation-specific killing of tumor cells [57]. Nair et al. employed an antigen-specific strategy by vaccinating mice with dendritic cells pulsed with MBT-2 tumor extracts [64]. They were able to demonstrate cytolytic activity in vitro and resistance to subsequent tumor rechallenge.

Several investigators are studying the transfer of genes to bladder cancer cells in order to establish a whole-tumor cell vaccine for treating localized and systemic disease. Larchian et al. described a tumor cell vaccine in an orthotopic murine model in which cells transfected with IL-2 or B7 genes were administered systemically [51]. This approach resulted in significant tumor regression and survival benefit, especially in mice treated with the combined vaccine (IL-2 and B7). Connor et al. has also successfully used IL-2 gene transfection as a cancer vaccine strategy for therapy in a murine model of bladder cell carcinoma [20]. Chen et al. used a recombinant adenoviral vector to transfer the IL-12 gene to the MB49 cell line and demonstrated significant tumor inhibition as well as subsequent specific protective immunity [17].

Several studies have also looked at the in vivo transduction of tumor cells to induce an immune response as an in situ tumor cell vaccine. Based on the easy access to the relatively nonabsorptive genitourinary epithelium as well as the vast experience with intravesical treatments for several bladder diseases, many consider bladder cancer to be an attractive candidate for such strategies. In this regard, Lee et al. [52] have demonstrated infectivity of murine and human transitional cell lines by a recombinant vaccinia vector that may be optimally suited for an immunotherapy protocol.

### **Conclusions**

Dramatic advances in available molecular techniques and growing understanding of antigen presentation and recognition have led to much renewed interest in immunotherapy as a possible adjuvant or alternative therapy for cancer. Those investigating such possibilities for urologic malignancies have been leaders in the field. Novel preclinical studies and early clinical trials have resulted in numerous successes; however, we should take care to interpret this data cautiously as tumor immunotherapy emerges from its infancy.

#### References

- Abbas AK, Murphy KM, Sher A (1996) Functional diversity of helper T lymphocytes. Nature 383: 787
- Adler A, Gillon G, Lurie H, Shaham J, Loven D, Shachter Y, Shani A, Servadio C, Stein JA (1987) Active specific immunotherapy of renal cell carcinoma patients: a prospective randomized study of hormomo-immuno- versus hormono-

- therapy. Preliminary report of immunological and clinical aspects. J Biol Respir Med 9: 610
- Alexander RB, Rosenberg SA (1990) Long-term survival of adoptively transferred tumor-infiltrating lymphocytes in mice. J Immunol 145: 1615
- Anton P, Tanke HJ, Allehoff EP, Kuczyk MA, Stief CG, Jonas U (1995) Localized renal cell carcinoma: detection of abnormal cells in peritumoral tissue. A cytophotometry and immunocytochemistry study. World J Urol 13: 149
- Asher AL, Mule JJ, Kasid A, Restifo NP, Salo JC, Reichert CM, Jaffe G, Fendly B, Kriegler M, Rosenberg SA (1993) Murine tumor cells transduced with the gene for tumor necrosis factor-alpha. Evidence for paracrine immune effects of tumor necrosis facto against. J Immunol 150: 1458
- Bander NH, Digvi C, Finn R, Larson S, Old LJ (1996) Renal cancer imaging with monoclonal antibody. J Urol 155: 583A
- Bander NH, Yao D, Liu H, Chen YT, Steiner M, Zuccaro W, Moy P (1997) MHC class I and II expression in prostate carcinoma and modulation by interferon-alpha and -gamma. Prostate 33: 233
- Baskar S, Ostrand-Rosenberg S, Nabavi N, Nadler L, Freeman G, Glimcher L (1993) Constitutive expression of B7 restores immunogenicity of tumor cells expressing truncated major histocompatability complex class II molecules. Proc Natl Acad Sci U S A 90: 5687
- Belldegrun A, Pierce W, Kaboo R, Tso CL, Turcillo P, Moldawer N, Golub S, de Kernion J, Figlin RA (1993) Interferon-alpha primed tumor infiltrating lymphocytes combined with interleukin-2 and interferon-alpha as therapy for metastatic renal cell carcinoma. J Urol 150: 1384
- Bennett MW, O'Connell J, O'Sullivan GC, Roche D, Brady C, Kelly J, Collins JK, Shanahan F (1999) Expression of Fas ligand by human gastric adenocarcinomas: a potential mechanism of immune escape in stomach cancer. Gut 44: 156
- Bjorkman PJ, Davis MM (1989) Model for the interaction of T cell receptors with the peptide/MHC complexes. Cold Spring Harb Symp Quant Biol 54: 365
- Bjorkman PJ, Saper MA, Samraoui B, Bennett WS. Strominger JL. Wiley DC (1987) Structure of the human class I histocompatability antigen, HLA-A2. Nature 329: 506
- Boon T, Cerottini J, Van den Eynde B, Van der Bruggen P, Van Pel A (1994) Tumor antigens recognized by T lymphocytes. Annu Rev Immunol12: 337
- Brown JH, Jardetsky TS, Gorga JC, Stern LJ, Urban RG, Strominger JL, Wiley DC (1993) The three-dimensional structure of the human class II histocompatability antigen, HLA-DR-1. Nature 364: 33
- Carrato C, Balague C, De Bolos C, Gonzalez E; Gambus G, Planas J, Perini JM, Andreu D, Real FX (1994) Differential apomucin expression in normal and neoplastic tissues. Gastroenterology 107: 160
- Cesano A, Visnneau S, Santoli D (1998) TALL-104 cell therapy of human solid tumors implanted in immunodeficient (SCID) mice. Anticancer Res 18: 2289
- 17. Chen L, Chen D, Block E, O'Donnell M, Kufe DW, Clinton SK (1997) Eradication of murine bladder carcinoma by intratumor injection of a bicistronic adenoviral vector carrying cDNAs for the IL-12 heterodimer and its inhibition by the IL-12 p40 subunit homodimer. J Immunol 159: 351
- Chen X, DeWolf WC, O'Donnell MA (1997) Synergistic effects of co-stimulatory cytokines on BCG-induced interferongamma production. J Urol 157: 1329A
   Childs R, Chernoff A, Contenin N, Bahceci E, Schrump D,
- Childs R, Chernoff A, Contenin N, Bahceci E, Schrump D, Leitman S, Read EJ, Tisdale J, Dunbar C, Linehan WM, Young NS, Barrett AJ (2000) Regression of metastatic renal-cell carcinoma after nonmyeloablative allogenic peripheral-blood stem-cell transplantation. N Engl J Med 343: 750
- Connor J, Bannerij R, Saito S, Heston W, Fair W, Gilboa E (1993) Regression of bladder tumors in mice treated with interleukin-2 gene-modified tumor cells. J Exp Med 177: 1127

- Correale P, Walmsley K, Zaremba S, Zhu M, Schlom J, Tsang KY (1998) Generation of human cytolytic T lymphocyte lines directed against prostate-specific antigen (PSA) employing a PSA oligoepitope peptide. J Immunol 161: 3186
- employing a PSA oligoepitope peptide. J Immunol 161: 3186
  22. Dranoff G, Soiffer R, Lynch T, Mihm M, Jung K, Kolesar K, Liebster L, Lam P, Duda R, Mentzer S, Singer S, Tanabe K, Johnson R, Sober A, Bhan A, Clift S, Cohen L, Parry G, Rokovich J, Richards L, Drayer J, Berns A, Mulligan RC (1997) A phase 1 study of vaccination with irradiated melanoma cells engineered to secrete human granulocyte-macrophage colony stimulating factor. Hum Gene Ther 8: 111
- 23. Dudley ME, Roopenian DC (1996) Loss of a unique tumor antigen by cytotoxic T lymphocyte immunoselection from a 3-methylcholanthrene-induced mouse sarcoma reveals secondary unique and shared antigens. J Exp Med 184: 441
- 24. Figlin RA, Pierce WC, Kaboo R, Tso CL, Moldawer N, Gitlitz B, deKernion J, Belldegrun A (1997) Treatment of metastatic renal cell carcinoma with nephrectomy, interleukin-2 and cytokine-primed or CD8(+) selected tumor infiltrating lymphocytes from primary tumor. J Urol 158: 740
- Finke JH, Zea AH, Stanley J, Longo DL, Mizoguchi H, Tubbs RR, Wiltrout RH, O'Shea JJ, Kudoh S, Klein E (1993) Loss of T cell receptor ζ chain and p56<sup>lck</sup> in T cells infiltrating human renal cell carcinoma. Cancer Res 53: 5613
- 26. Fu YX, Watson GA, Kassahara M, Lopez DM (1991) The role of tumor-derived cytokines on the immune system of mice bearing a mammary adenocarcinoma. I. Induction of regulatory macrophages in normal mice by the in vivo administration of rGM-CSF. J Immunol 146: 783
- 27. Galligioni E, Quaia M, Merlo A, Carbone A, Spada A, Favaro D, Santarosa M, Sacco C, Talamini R (1996) Adjuvant immunotherapy treatment of renal carcinoma patients with autologous tumor cells and bacillus Calmette-Guerin. Cancer 77: 2560
- Gambus G, Bolos CD, Andreu D, Franci C, Egea G, Real FX (1993) Detection of the MUC2 apomucin tandem repeat with a mouse monoclonal antibody. Gastroenterology. 104: 93
- Gansbacher B, Houghton, Livingston P (1992) A pilot study of immunization with HLA-A2 matched allogeneic melanoma cells that secrete interleukin-2 in patients with metastatic melanoma. Hum Gene Ther 3: 677
- 30. Gansbacher B, Motzer R, Houghton A (1992) A pilot study of immunization with interleukin-2 secreting allogeneic HLA-A2 matched renal cell carcinoma cells in patients with advanced renal cell carcinoma. Hum Gene Ther 3: 691
- 31. Golumbek PT, Lazenby A, Levitsky HI, Jaffee LM, Karasuyama H, Baker M, Pardoll DM (1991) Treatment of established renal cancer by tumor cells engineered to secrete interleukin-4. Science 254: 713
- 32. Goodfield J (1984) Dr. Coley's Toxins. Science 226: 68
- 33. Grabamaier K, Vissers JL, De Weijert MC, Oosterwijk-wakka JC, Van Bokhoven A, Brakenhoff RH, Noessner E, Mulders PA, Merkx G, Figdor CG (2000) Molecular cloning and immunogenicity of renal cell carcinoma-associated antigen G250. Int J Cancer 85: 865
- Greten TF, Jaffee EM (1999) Cancer vaccines. J Clin Oncol 17: 1047
- 35. Haynes B, Denning SM, Le PT, Singer KH (1990) Human intrathymic T cell differentiation. Semin Immunol 2: 67
- 36. Healy CG, Simons JW, Carducci MA, DeWeese TL, Bart-kowski M, Tong KP, Bolton WE (1998) Impaired expression and function of signal-transducing zeta chains in peripheral T cells and natural killer cells in patients with prostate cancer. Cytometry 32: 109
- 37. Myers RB, Srivastava S, Grizzle WE (1995) Lewis Y antigen as detected by the monoclonal antibody BR96 is expressed strongly in prostatic adenocarcinoma. J Urol 153: 1572
- 38. Hsu FJ, Benike C, Fagoni F, Liles TM, Czerwinski D, Taidi B, Engleman EG, Levy R (1996) Vaccination of patients with B-cell lymphoma using autologous antigen-pulsed dendritic cells. Nat Med 2: 52

- 39. Irvine K, McCabe B, Rosenberg S, Restifo NP (1995) Synthetic oligonucleotide expressed by a recombinant vaccinia virus elicits therapeutic CTL. J Immunol 154: 4651
- Isreaeli RS, Powell T, Corr JG, Fair WR, Heston WD (1994)
   Expression of prostate-specific membrane antigen. Cancer Res 54: 1807
- 41. Julliard V, Villefroy P, Godfrin D, Pavirani A, Venet A, Guillet JG (1995) Long-term humoral and cellular immunity induced by a single immunization with replication-defective adenovirus recombinant vector. Eur J Immunol 25: 3467
- 42. June CH, Bluestone JA, Nadler LM, Thompson CB (1994) The B7 and CD28 receptor families. Immunol Today 11: 191
- 43. Kass E, Schlom J, Thompson J, Guadagni F, Graziano P, Greiner JW (1999) Induction of protective host immunity to carcinoembryonic antigen (CEA), a self antigen in CEA transgenic mice, by immunizing with a recombinant vaccinia-CEA virus. Cancer Res 59: 676
- 44. Kawakita M, Rao GS, Ritchey JK, Ornstein DK, Hudson MA, Tartaglia J, Paoletti E, Humphrey PA, Harmon TJ, Ratliff TL (1997) Effect of canarypox virus (ALVAC)-mediated cytokine expression on murine prostate tumor growth. J Natl Cancer Inst 89: 428
- 45. Kiamura K, Livingston PO, Fortunato SF (1995) Serological response patterns of melanoma patients immunized with a GM2 ganglioside conjugate vaccine. Proc Natl Acad Sci U S A 92: 2805
- 46. Kim J, Park TK, Hu S (1995) Defining the molecular recognition of GLOBO H (human breast cancer) antigen through probe structures prepared by total synthesis. J Org Chem 60: 7716
- 47. Kim YS, Kim KH, Choi JA, Lee JH, Kim HK, Won NH, Kim I (2000) Fas (APO-1/CD95) ligand and Fas expression in renal cell carcinomas: correlation with the prognostic factors. Arch Pathol Lab Med 124: 687
- 48. Kuriyama S, Tominaga K, Kikukawa M, Nakatani T, Tsujinoue H, Yamazaki M, Nagao S, Toyokawa Y, Mitoro A, Fukui H (1998) Inhibitory effects of human sera on adenovirus-mediated gene transfer into rat liver. Anticancer Res 18: 2345
- 49. Lahat N, Alexander N, Levin DR, Moskowitz B (1989) The relationship between clinical stage, natural killer activity and related immunological parameters in adenocarcinoma of the prostate. Cancer Immunol Immunother 28: 208
- Lamm DL, Blumenstein BA, Crawford ED, Montie JE, Scardino P, Grossman HB, Stanisic TH, Smith JA, Sullivan J, Sarosdy MF (1991) A randomized trial of intravesical doxorubicin and immunotherapy with bacille Calmette-Guerin for transitional-cell carcinoma of the bladder. N Engl J Med 325: 1205
- Larachian WA, Robertson K, Robertson C, Gilboa E, Heston WDW, Fair WR (1997) Liposome mediated gene transfer in human bladder cancer cells. J Urol 157: 1196A
- Lee SS, Eisenlohr LC, McCue PA, Mastrangelo MJ, Lattime EC (1994) Intravesical gene therapy: in vivo gene transfer using recombinant vaccinia virus vectors. Cancer Res 54: 3325
- Lehmann C, Zeis M, Schmitz N, Uharek L (2000) Impaired binding of perforin on the surface of tumor cells is a cause of target cell resistance against cytotoxic effector cells. Blood 96: 594
- Lenschow DJ, Walunas TL, Bluestone JA (1996) CD28/B7 system of T cell costimulation. Annu Rev Immunol 14: 233
- 55. Levin, I, Klein T, Kuperman O, Segal S, Shapira J, Gal R, Hart Y, Klein B (1994) The expression of HLA class I antigen in prostate cancer in relation to tumor differentiation and patient survival. Cancer Detect Prevent 18: 443
- Lubaroff DM, Cohen MB, Schultz LD, Beamer WG (1995) Survival of human prostate carcinoma, benign hyperplastic tissues, and IL-2-activated lymphocytes in scid mice. Prostate 27: 32

- Luo Y, Chen X, Han R, Chorev M, Dewolf WC, O'Donnell MA (1999) Mutated ras p21 as a target for cancer therapy in mouse transitional cell carcinoma. J Urol 162: 1519
- 58. Marchand M, Weynants P, Rankin E, Arienti F, Belli F, Parmiani G, Cascinelli N, Bourland A, Vanwijck R, Humblet Y, Canon JL, Laurent C, Brasseur F, Herman J, Coulie P, Boon T (1995) Tumor regression responses in melanoma patients treated with a peptide encoded by gene MAGE-3. Int J Cancer 63: 883
- McCune CS, Schapira DV, Mastrangelo MJ (1981) Specific immunotherapy of advanced renal carcinoma: evidence for the polyclonality of metastases. Cancer 27: 1984
- 60. McLaughlin JP, Schlom J, Kantor JA, Greiner JW (1996) Improved immunotherapy of a recombinant carcinoembryonic antigen vaccinia vaccine when given in combination with interleukin-2. Cancer Res 56: 2361
- 61. Montie JE, Stewart BH, Straffon RA, Banowsky LH, Hewitt CB, Montague DK (1977) The role of adjunctive nephrectomy in patients with metastatic renal cell carcinoma. J Urol 117: 272
- Morales A, Eidinger D, Bruce AW (1976) Intracavitary bacillus Calmette Guerin in the treatment of superficial bladder tumors. J Urol 116: 180
- 63. Mullbacher A, Bellett A, Hla R (1989) The murine cellular immune response to adenovirus type 5. Immunol Cell Biol 67:
- 64. Nair SK, Snyder D, Rouse BT, Gilboa E (1997) Regression of tumors in mice vaccinated with professional antigen-presenting cells with tumor extracts. Int J Cancer 70: 706
- Nestle FO, Alijagic S, Gilliet M, Sun Y, Grabbe S, Dummer R, Burg G, Schadendorf D (1998) Vaccination of melanoma patients with peptide- or tumor lysate-pulsed dendritic cells. Nat Med 4: 328
- 66. Norman A, Parker S, Lew D, Manthrope M, Marquet M (1995) Preclinical pharmacokinetics, manufacturing and safety studies supporting a multicenter cancer gene therapy trial. Hum Gene Ther 6: 549
- 67. O'Donnell MA, Duda RB, Yang U (1993) Construction of interleukin-2 secreting BCG. J Urol 149: 228A
- Orstrand-Rosenberg S, Thakur A, Clements V (1983) Rejection of mouse sarcoma cells after transfection of MHC class II genes. J Exp Med 144: 4068
- 69. Pan ZK, Ikonomidis G, Lazenby A, Pardoll D, Paterson Y (1995) A recombinant Listeria Monocytogenes vaccine expressing a model tumor antigen protects mice against lethal tumor cell challenge and causes regression of established tumors. Nat Med 1: 471
- Pardoll DM (1994) Tumor antigens: a new look for the 1990s. Nature 369: 357
- Parkhurst MR, Salgaller ML, Southwood S, Robbins PF, Sette A, Rosenberg SA, Kawakami Y (1996) Improved induction of melanoma-reactive CTL with peptides from the melanoma antigen gp100 modified at HLA-A\*0201-binding residues. J Immunol 157: 2539
- Prehn RT, Main JM (1957) Immunity to methylcholanthrene induced sarcomas. J Natl Cancer Inst 18: 769
- 73. Putzer BM, Hitt M, Mueller WJ, Emtage P, Gauldie J, Graham FL (1997) Interleukin-12 and B7-1 costimulatory molecule expressed by an adenoviral vector act synergistically to facilitate tumor regression. Proc Natl Acad Sci U S A 94: 10889
- Radjoa S, Frey AB (2000) Cancer-induced defective cytotoxic
   T lymphocyte effector function: another mechanism how antigenic tumors escape immune-mediated killing. Mol Med
   465
- Ratliff TL, Gillen D, Catalona WJ (1987) Requirement of a thymus-dependent immune response for BCG-mediated antitumor activity. J Urol 137: 15
- Ratliff TL, Ritchey IK, Yuan JJ, Andriole GL, Catalona WJ (1993) T cell subsets required for intravesical BCG immunotherapy for bladder cancer. J Urol 150: 1018

- 77. Rauschmeier HA (1988) Immunotherapy of metastatic renal cancer. Semin Surg Oncol 4: 169
- Restifo NP, Bacik I, Irvine KR, Yewdell JW, McCabe BJ, Anderson RW, Eisenlohr LC, Rosenberg SA, Bennink JR (1995) Antigen processing in vivo and the elicitation of primary CTL responses. J Immunol 154: 4414
- Rosenberg SA (1996) Development of cancer immunotherapies based on identification of the genes encoding cancer regression antigens. J Natl Cancer Inst 88: 1635
- 80. Rosenberg SA, Spiess P, Lafreniere R (1986) A new approach to adoptive immunotherapy of cancer with tumor-infiltrating lymphocytes. Science 233: 1318
- Rosenberg SA, Lotze MT, Yang JC (1993) Prospective randomized trial of high dose interleukin-2 alone or in combination with lymphokine-activated killer cells for the treatment of patients with advanced cancer. J Natl Cancer Inst 85: 622
- Rosenberg SA, Yang JC, Topalian SL, Schwartzentruber HJ, Weber JS, Parkinson DR, Seipp CA, Einhorn JH, White DE (1994) Treatment of 283 consecutive patients with metastatic melanoma or renal cell carcinoma using high dose bolus interleukin-2. JAMA 271: 907
- 83. Rosenberg SA, Yang JC, Schwartzentruber DJ, Hwu P, Marincola FM, Topalian SL, Restifo NP, Dudley ME, Schwarz SL, Spiess PJ, Wunderlich JR, Parkhurst MR, Kawakami Y, Seipp CA, Einhorn JH, White DE (1998) Immunologic and therapeutic evaluation of a synthetic peptide vaccine for the treatment of patients with metastatic melanoma. Nat Med: 321
- 84. Rosenberg SA, Zhai Y, Yang JC, Schwartzentruber DJ, Hwu P, Marincola FM, Topalian SL, Restifo NP, Seipp CA, Einhorn JH, Roberts B, White DE (1998) Immunizing patients with metastatic melanoma using recombinant adenoviruses encoding MART-1 or gp100 melanoma antigens. J Natl Cancer Inst 90: 1894
- Sahasrabudhe DM, DeKernion JB, Pontes JE, Ryan DM, O'Donnell RW, Marquis DM, Mudholkar GS, McCune CS (1986) Specific immunotherapy with suppressor function inhibition for metastatic renal cell carcinoma. J Biol Respir Med 5: 581
- Salgaller ML, Tjoa BA, Lodge PA, Ragde H, Kenny G, Boynton A, Murphy GP (1998) Dendritic cell-based immunotherapy of prostate cancer. Crit Rev Immunother 18: 109
- 87. Sanda MG, Ayyagari SR, Jeffer EM, Epstein FI, Clift SL, Cohey LK, Dranoff G, Pardoll MD, Mulligan RC, Simons JW (1994) Demonstration of a rational strategy for human prostate cancer gene therapy. J Urol 151: 622
- prostate cancer gene therapy. J Urol 151: 622 88. Santori A, Palmeieri G, Procopio A (1989) Mechanism of target cell killing by natural killer cells. Ital J Med 4: 59
- 89. Sarna G, Figlin R, DeKernion JB (1987) Interferon in renal cell carcinoma. Cancer 59: 610
- Seigler HF, Darrow TL, Abdel-wahab Z, Gangavalli R, Barber J (1994) A phase 1 trial of human gamma interferon transduced autologous tumor cells in patients with disseminated malignant melanoma. Hum Gene Ther 5: 761
- 91. Shu S, Chou T, Sakai K (1989) Lymphocytes generated by in vivo priming and in vitro sensitization demonstrate therapeutic efficacy against a murine tumor that lacks apparent immunogenicity. J Immunol 143: 740
- 92. Simons JW, Jaffee EM, Weber CE, Levitsky HI, Nelson WG, Carducci MA, Lazenby AJ, Cohen LK, Finn CC, Clift SM, Hauda KM, Beck LA, Leiferman KM, Owens AH Jr, Piantadosi S, Dranoff G, Mulligan RC, Pardoll DM, Marshall FF (1997) Bioactivity of autologous irradiated renal cell carcinoma vaccines generated by ex vivo granulocyte-macrophage colony-stimulating factor gene transfer. Cancer Res 57: 1537
- Slovin SF, Livingston PO, Rosen N, Sepp-Lorenzino L, Kelly WK, Mendelsohn J, Scher HI (1996) Targeted therapy for prostate cancer: the Memorial Sloan-Kettering Cancer Center approach. Semin Oncol 23: 41
- 94. Specht JM, Wang G, Do MT, Lam JS, Royal RE, Reeves ME, Rosenberg SA, Hwu P (1997) Dendritic cells retrovirally

- transduced with a model tumor antigen gene are therapeutically effective against established pulmonary metastases. J Exp Med 186: 1213
- 95. Spiess PJ, Yang JC, Rosenberg SA (1987) Tumor-specific cytolysis by lymphocytes infiltrating lymphocytes expanded in recombinant interleukin-2. J Natl Cancer Inst 79: 1067
- 96. Tang DC, DeVit M, Johnston SA (1992) Genetic immunization is a simple method for eliciting an immune response. Nature 356: 152
- 97. Tjoa B, Boynton A, Kenny G, Radge H, Misrock SL, Murphy G (1995) Presentation of prostate tumor antigens by dendritic cells stimulates T-cell proliferation and cytotoxicity.
- 98. Tjoa BA, Simmons SJ, Elgamal A, Rogers A, Radge H, Kenny GM, Troychak MJ, Boynton AL, Murphy GP (1999) Follow-up evaluation of a phase II prostate cancer vaccine trial. Prostate 40: 125
- 99. Townsend SE, Allison JP (1993) Tumor rejection after direct costimulation of CD8+ T cells by B7-transfected melanoma cells. Science 259: 368
- 100. Trowsdale J, Tagoussis J, Campbell RD (1991) Map of the human MHC. Immunol Today 12: 443
- 101. van Dijk J, Uemura H, Beniers AJ, Peelen WP, Zegveld ST, Fleuren GJ, Warnaar SO, Oosterwijk E (1994) Therapeutic effects of monoclonal antibody G250, interferons and tumor necrosis factor, in mice with renal cell carcinoma xenografts. Int J Cancer 56: 262
- 102. Vieweg J, Rosenthal FM, Bannerji R, Heston WDW, Fair WR, Gansbacher B, Gilboa E (1994) Immunotherapy of prostate cancer in the dunning rat model: use of cytokine gene modified tumor vaccines. Cancer Res 54: 1760
- 103. Vissers J, De Vries IJ, Schreurs MW, Engelen LP, Oosterwijk E, Figdor CG, Adema GJ (1999) The renal cell carcinoma-associated antigen G250 encodes a human leukocyte antigen (HLA)-A2.1-restricted epitope recognized by cytotoxic T lymphocytes. Cancer Res 59: 5554
- 104. Von der Maase H, Geertsen P, Thatcher N, Jasmin C, Mercatello A, Fossa SD, Symann M, Stoter G, Nagel G, Israel L (1991) Recombinant interleukin-2 in metastatic renal cell carcinoma a European multicenter phase-II study. Eur J Cancer 27: 1583
- 105. Wallich R, Bulbuc N, Hammerling GJ, Katzav S, Segal S, Feldman M (1985) Abrogation of metastatic properties of tumor cells by de novo expression of H-2 K antigens following H-2 gene transfection. Nature 315: 301
- 106. Weitjens ME, Hart EH, Bolhuis RL (1999) Phase 1 radioimmunotherapy of metastatic renal cell carcinoma with 131Ilabelled chimeric monoclonal antibody G250. Clin Cancer Res 5 [Suppl 10]: 3268s
- Wirth MP (1993) Immunotherapy for metastatic renal cell carcinoma. Urol Clin N Am 20: 283
- 108. Xiang R, Lode HN, Chao TH, Ruehlmann JM, Dolman CS, Rodriguez F, Whitton JL, Overwijk WW, Restifo NP, Reisfeld RA (2000) An autologous oral DNA vaccine protects against murine melanoma. Proc Natl Acad Sci U S A 97: 5492
- 109. Yang Y, Ertl HC, Wilson JM (1994) MHC class I-restricted cytotoxic T lymphocytes to viral antigens destroy hepatocytes in mice infected with E1-deleted recombinant adenoviruses. Immunity 1: 433
- Yoshimura I, Heston WDW, Gansbacher B, Fair WR (1996)
   Cytokine mediated immuno-gene therapy in rat prostate cancer model. J Urol 155: 798
- 111. Zea A, Curti B, Longo D, Alvord W, Strobl SL, Mizoguchi H, Creekmore SP, O'Shea JJ, Powers GC, Urba WJ (1995) Alterations in T cell receptor and signal transduction molecules in melanoma patients. Clin Cancer Res 1: 1327
- 112. Zhai Y, Yang JC, Kawakami Y, Spiess P, Wadsworth SC, Cardoza LM, Couture LA, Smith AE, Rosenberg SA (1996) Antigen-specific tumor vaccines: development and characterization of recombinant adenoviruses encoding MART1 or gp100 for cancer. J Immunol 1556: 700